

MCNAIR SCHOLARS PROGRAM

The Future of Biodiesel & Wastewater Treatment **Brissa Acevedo and Dr. Matthew L. Alexander** Wayne H. King Department of Chemical and Natural Gas Engineering

ABSTRACT

Microalgae are an excellent vehicle for significant triglyceride production, which can be transformed chemically into biodiesel fuel. However, large volumes of water are necessary for successful microalgae cultivation. With ever-increasing demands for clean potable water, it makes sense to investigate using non-potable waters, such as wastewater and brackish waters, for microalgae culturing. The current research aim is to investigate whether the microalgae strains Dunaliella salina, and Neochloris oleoabundans are good candidates for industrial triglyceride production and treatment of wastewater by comparing their growth in the basic media versus growth in wastewater media that has additional sodium nitrate (NaNO₃) and sodium sodium phosphate dibasic (Na₂HPO₄) or dipotassium phosphate (K_2HPO_4) (depending on the corresponding media). The expected results are that the wastewater media will have a higher microalgae production and thus oil production than that of the basic media. With this in mind, microalgae might be a great candidate for industrial feedstock production and treatment of wastewater.

INTRODUCTION

Algae:

- photosynthetic organisms that grow in a range of aquatic habitats, such as lakes, ponds, rivers, oceans, and even wastewater
- Qualities:
- Capacity to survive harsh environmental conditions (Ajala & Alexander, 2020a) Produce Lipids
- Triglycerides (TAG)
- Great candidate to potentially replace fossil fuels (Hibel et al., 2015; Mondal et al., 2017; Morteo & Alexander, 2022; Patil et al., 2008)
- Resource to potable water (Ajala & Alexander, 2020; Mondal et al., 2017)

PURPOSE OF THE STUDY

The current research's aim is to investigate if the microalgae strains, D. salina and N. oleoabundans are good candidates for industrial feedstock production and treatment of wastewater by comparing them to the medium that does not alter the concentrations of sodium nitrate (NaNO₃) and sodium phosphate dibasic (Na₂HPO₄) or dipotassium phosphate (K_2 HPO₄) (depending on the corresponding media).

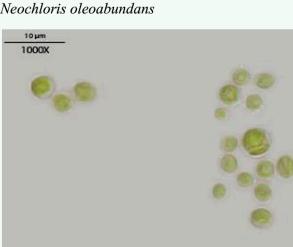
RESEARCH QUESTIONS

- Does a medium in higher sodium nitrate $(NaNO_3)$ and sodium phosphate dibasic (Na_2HPO_4) or dipotassium phosphate (K_2HPO_4) (depending on the corresponding media) concentrations produce more triglycerides than that of the algae harvested in the non-altered medium?
- Is there a correlation between algae growth and triglycerides (oil production)?
- How does the growth of algae affect the treatment of wastewater and the overall release of triglycerides?

METHODS

The following procedures will be performed to investigate if the microalgae strains, Dunaliella salina, and Neochloris oleoabundans are good candidates for industrial feedstock production and treatment of wastewater by comparing it to the medium that does not alter the concentrations of sodium nitrate (NaNO₃) and sodium phosphate dibasic (Na_2HPO_4) or dipotassium phosphate (K_2HPO_4) (depending on the corresponding media). Figure 1





JTEX LB #1185 Neochloris oleoabundans Note. Purchased from UT-Austin.

Note. Purchased from UT-Austin

RESULTS

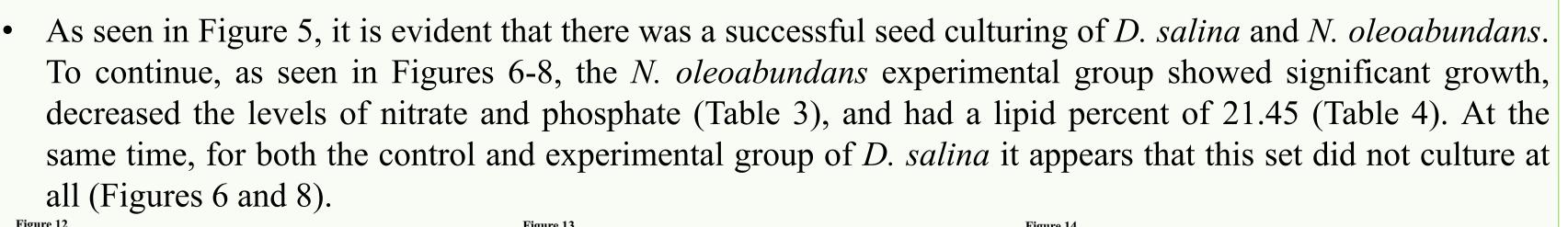


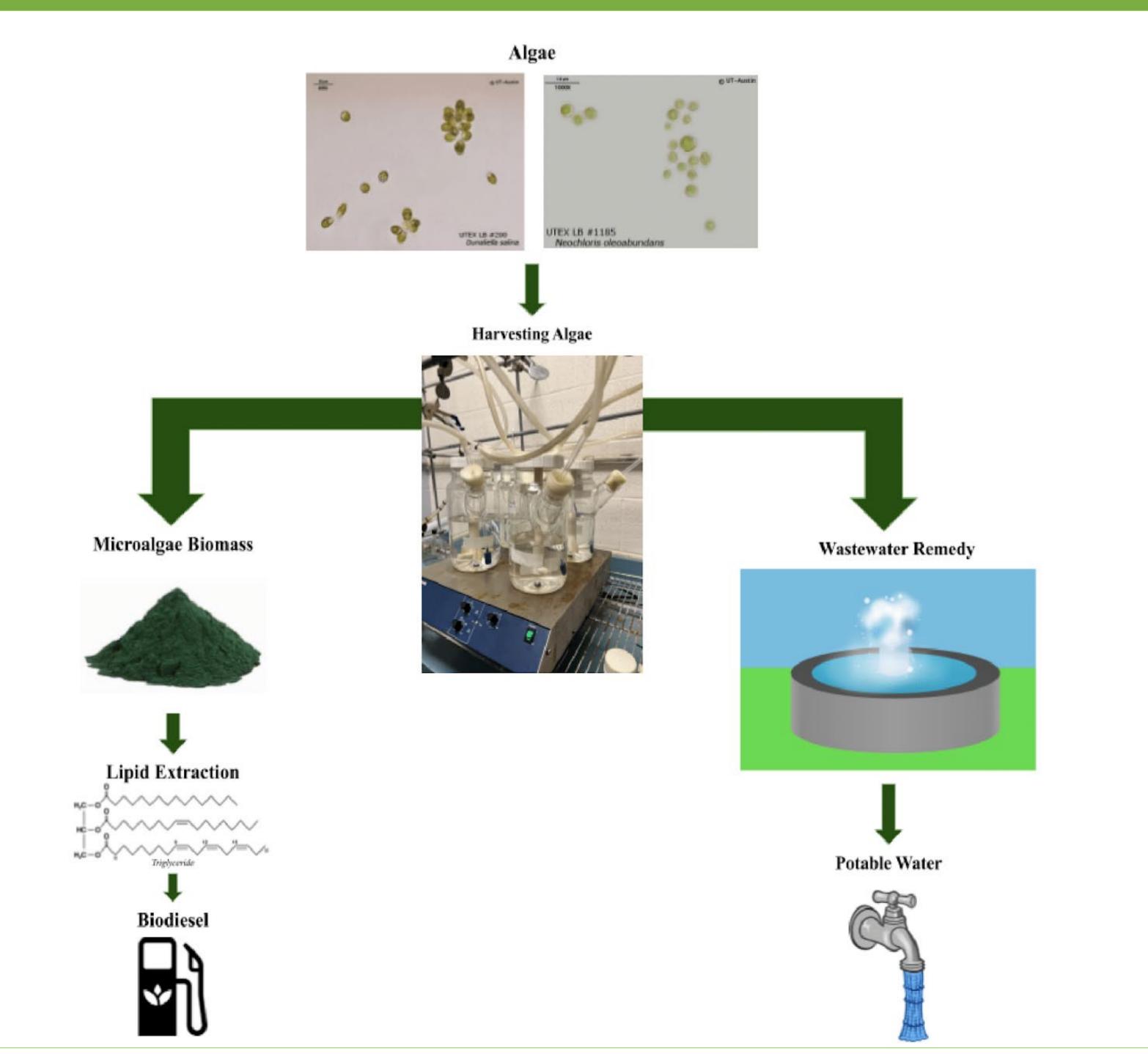
Figure 12		Figure 13	I	Figure 14		
Algae Growth: OD readings at 640 nm		Nitrate readings at 500 nm	Nitrate readings at 500 nm		Phosphate readings at 430 nm	
Algae Growth vs. ⁻	Time	Nitrate Levels vs. Time		Phosphate Levels vs. Time		
1.00 United Density at 640 m 0.75 0.20 0.20 0.20 0.20 0.20 0.20 0.20 0.2	 D. salina Experimental (OD) - N. oleoabundans N. oleoabundans Experimental (OD) 1000 1	Control (OD) D. salina Control (mg/L) N. oleoabundans E 20.0 15.0 10.0 5.0 0.0 7/10/2022 7/17/2022 7/24/202	Experimental (mg/L)	 D. salina Control (mg/L) D. salina Experimental N. oleoabundans Experimental Solo 40.0 40.0		
Note. OD reading over 3 wee		<i>Note.</i> Nitrate level readings over 3 weeks of Biore Spectrophotometer.		<i>Note.</i> Phosphate level readings over 3 weeks of Biore Spectrophotometer.	eactors 1-4 using the Hach DR/2000 Visi	
Figure 6	Figure 7	Table 3	Ta	able 4		
Bioreactors 1 and 2: D	D. salina Bioreactors 3 and 4: N. oleod	bundans				
	Star of	Biomass Per Dry Weight	Lij	pid Percent		
	al.	Bion	Biomass (gwd)		pid %	
		N. oleoabundans Control	0.04574285714	N. oleoabundans Control	5	

Note. Bioreactor 1 (front): D. salina control;

Biomass (gwd)		
N. oleoabundans Control		
N. oleoabundans Experimental		
D. salina Control (seed culture)		
<i>N. oleoabundans</i> Control (seed culture)		

Bioreactor 3 (front): *N. oleoabundans* control; Bioreactor 4 (back): *N. oleoabundans* experimental. *Note.* Biomass measured in grams.

CONCLUSION



Note. The lipid percent from the biomass.

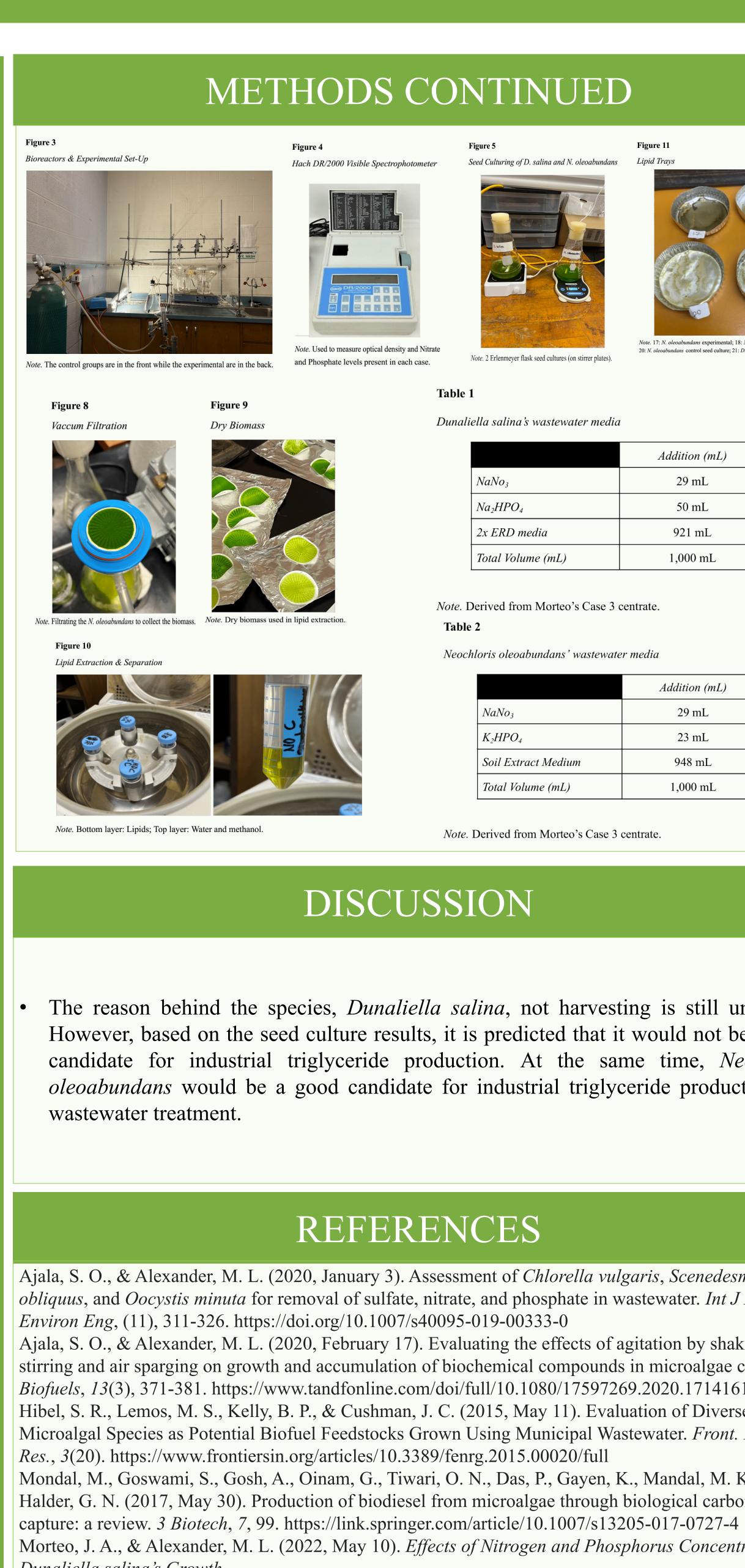
N. oleoabundans Experimental

D. salina Control (seed culture)

N. oleoabundans Control (seed

0.17242857

0.592



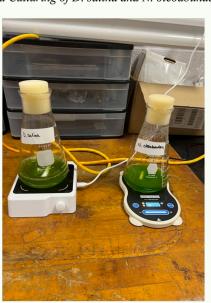
Dunaliella salina's Growth.

Dr. Matthew L. Alexander McNair Staff at Texas A&M University – Kingsville



METHODS CONTINUED

Seed Culturing of D. salina and N. oleoabundans



Note. 2 Erlenmeyer flask seed cultures (on stirrer plate



Table 1

Dunaliella salina's wastewater media

	Addition (mL)
NaNo ₃	29 mL
Na_2HPO_4	50 mL
2x ERD media	921 mL
Total Volume (mL)	1,000 mL

Note. Derived from Morteo's Case 3 centrate.

Table 2

Neochloris oleoabundans' wastewater media

	Addition (mL)	
NaNo ₃	29 mL	
K_2HPO_4	23 mL	
Soil Extract Medium	948 mL	
Total Volume (mL)	1,000 mL	

Note. Derived from Morteo's Case 3 centrate.

DISCUSSION

• The reason behind the species, *Dunaliella salina*, not harvesting is still unknown. However, based on the seed culture results, it is predicted that it would not be a good candidate for industrial triglyceride production. At the same time, Neochloris oleoabundans would be a good candidate for industrial triglyceride production and

REFERENCES

Ajala, S. O., & Alexander, M. L. (2020, January 3). Assessment of Chlorella vulgaris, Scenedesmus obliquus, and Oocystis minuta for removal of sulfate, nitrate, and phosphate in wastewater. Int J Energy

Ajala, S. O., & Alexander, M. L. (2020, February 17). Evaluating the effects of agitation by shaking, stirring and air sparging on growth and accumulation of biochemical compounds in microalgae cells. *Biofuels*, *13*(3), 371-381. https://www.tandfonline.com/doi/full/10.1080/17597269.2020.1714161 Hibel, S. R., Lemos, M. S., Kelly, B. P., & Cushman, J. C. (2015, May 11). Evaluation of Diverse Microalgal Species as Potential Biofuel Feedstocks Grown Using Municipal Wastewater. Front. Energy *Res.*, *3*(20). https://www.frontiersin.org/articles/10.3389/fenrg.2015.00020/full

Mondal, M., Goswami, S., Gosh, A., Oinam, G., Tiwari, O. N., Das, P., Gayen, K., Mandal, M. K., & Halder, G. N. (2017, May 30). Production of biodiesel from microalgae through biological carbon

Morteo, J. A., & Alexander, M. L. (2022, May 10). Effects of Nitrogen and Phosphorus Concentrations in

Patil, V., Tran, K. -Q., & Giselrod, H. R. (2008, July 9). Towards Sustainable Production of Biofuels from Microalgae. Int. J. Mol. Sci, 9(7), 1188-1195. https://doi.org/10.3390/ijms9071188

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